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## Insecticides Exposure during Early Life Alters Prostate Cells Differentiation in Adulthood Spraque Dawley Rat

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**Abstract:** Exposure of endocrine disruptor chemicals during early life may alter prostate morphogenesis and cellular differentiation, associated with an increased risk of prostatic intraepithelial neoplasia (PIN). This study was designed to elucidate the effect of early insecticide exposure in the prostate gland. Newborn male Spraque Dawley rats were allocated into untreated (control) and treated groups, including estrogen potent (25  $\mu$ g  $\beta$  estradiol 3-benzoate), mosquito coil repellent, and liquid mosquito repellent. Prostate luminal epithelial and periductal stromal cells were significantly altered among mosquito insecticide groups [mosquito coil repellent (p < 0.001), 3-ml (p < 0.002), and 4-ml liquid mosquito repellent (p < 0.009)] and [mosquito coil repellent (p < 0.001), 3-ml (p < 0.05) and 4-ml liquid mosquito repellent (p = 0.05)] compared to those of control group, respectively. Mosquito insecticide exposure during early life leads to an alteration of prostate cellular differentiation in adulthood that may predispose to prostate cancer.

Keywords: early life exposure, endocrine disruptor chemicals, insecticides exposure, mosquito repellent, prostate cancer.

## 生命早期的杀虫剂暴露会改变成年斯普拉格·道利大鼠的前列腺细胞分化

摘要:早期暴露于内分泌干扰物的化学物质可能会改变前列腺形态发生和细胞分化,从 而增加前列腺上皮内瘤变(别针)的风险。该研究旨在阐明早期杀虫剂在前列腺中的暴露作 用。将新生的斯普拉格·道利雄性大鼠分为未治疗组(对照组)和治疗组,包括雌激素有效( 25微克雌二醇3-

苯甲酸酯),驱蚊剂和驱蚊剂。蚊虫杀虫剂组之间的前列腺腔上皮和导管周间质细胞发生了 显着变化[蚊香驱蚊剂(p <0.001),3-毫升(p <0.002)和4-ml液体蚊香剂(p <0.009)]和[蚊香驱蚊剂(p <0.001),3毫升(p <0.05)和4毫升液体蚊香(p = 0.05)]。生命早期暴露于蚊虫杀虫剂会导致成年期前列腺细胞分化的改变,这可能导致前列 腺癌。

关键字:早期生活暴露,内分泌干扰物化学物质,杀虫剂暴露,驱蚊剂,前列腺癌。

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## **1. Introduction**

Over the past decades, there has been increasing concern on the impact of environmental contaminants exposure known as endocrine disruptor chemicals (EDCs), chemicals that may interfere with the biosynthesis, metabolism, and/or action of endogenous hormones. Alterations of male reproductive development and health that have been reported to be associated with EDCs range from testicular dysgenesis syndrome (TDS), which includes demasculinization or feminization, cryptorchidism, hypospadias, in situ germ cell testicular carcinoma, reduced semen quality, and progression of prostate cancer [1], [2], [3].

Prostate cancer is the most common malignancy in men and the second cause of cancer deaths in developed countries. In the United States, the incidence (1.6-fold) and mortality rate (2.4-fold) among the African-American population were reported higher compared to the white population [4]. Gene polymorphisms, environmental factors (EDCs), lifestyle, and endogenous factors (age, race, and family history) have been studied to comprehend the discrepancy. Prins S reported increasing evidence both from epidemiology studies and animal models determining the role of EDCs that may interfere with the development and progression of prostate cancer [5]. In humans, common agricultural pesticide exposure is associated with an increased rate of prostate cancer in men with a positive family history, suggesting geneticenvironmental interactions [6]. An increased risk of prostate cancer was reported in numerous studies related to populations exposed to long-term and lowinsecticides (organophosphate dose and organochlorine) [7]. Testosterone plays the primary role in prostate cancer; however, early-life exposure to estrogenic compounds, including EDCs, may also enhance prostate susceptibility to hormonal carcinogens in adulthood [8], [9].

Every tissue has a specific window of development, which refers to the critical period when the developing tissue is susceptible to the effects of EDC exposure [10]. The critical window of reproductive tract development is between 7 and 40 weeks of gestation, during which cells undergo rapid mitotic division and differentiation, eventually resulting in puberty and maturation. The exposure of EDCs during the critical window period will affect cell reprogramming [11]. Therefore, besides the dose and potency of EDCs, the timing of exposure is also very critical, as exposure during the window of development likely leads to irreversible effects. In contrast, the effects of exposure outside of the window of development can be reversed or mitigated with the removal of exposure [12]. Additionally, EDCs were designed to have a longer half-life as an industrial use benefit. Still, their longevity is detrimental to wildlife and humans, and some chemicals that were banned decades ago remain at high levels in the environment.

Tropical countries such as Indonesia have been affected more rigorously by infectious diseases than countries temperate due to biological and environmental factors. Mosquitoes are the most common transmitters of vector-borne human diseases in tropical countries due to environmental conditions (temperature and humidity) conducive for In addition, human cultural behavior reproduction. also plays a role in disease-transmission control programming. Children under the age of 5 have a higher risk of mosquito-borne diseases. Accordingly, to protect children from mosquito bites, mosquito repellent usage is prevalent, often without considering potential side effects [13], [14]. This study was aimed to elucidate the effect of early life insecticide exposure on prostate gland alteration in rats.

## 2. Methodology

### 2.1. Animal and Housing

A posttest-only control group was designed to assure the objectivity of the study. Male one-day-old Sparque Dawley (SD) rats were acclimatized for 2 days in standard cages under standard conditions (room temperature at  $26 \pm 3^\circ$ , 12-hour light/ dark cycle) and kept in standard cages at the Animal Care Unit in Gadjah Mada University, Yogyakarta. The rats were randomly selected and allocated into 5 groups: control (untreated), treated groups including 25  $\mu$ g  $\beta$  estradiol 3-benzoate, and insecticide groups [spiral mosquito coil repellent contains transfluthrin 0.03%/coil; 3-ml 4-ml mosquito liquid repellent contains and transfluthrin 0.162 mg and propoxur 4.05 mg/ml]. After weaning, the rats were housed up to 5 per cage. Drinking water was available in plastic bottles, and the rats were fed a pellet diet ad libitum from weaning to postnatal day (PND) 100. Pesticide exposure was designed to resemble the natural setting of mosquito repellent usage.

# 2.2. Administration of $\boldsymbol{\beta}$ Estradiol 3-Benzoate and Mosquito Repellent

Twenty-five  $\mu g$  of  $\beta$  estradiol 3-benzoate was diluted in 0.02 ml of sesame oil and administered subcutis in single doses with a non-traumatic needle during alternate days for 20 days. The exposure of mosquito insecticide was performed in a different room out of proximity to prevent effect interference. Multilevel cages were used to expose the rats to mosquito coil repellant fumes, which last up to 8 hours. Coils were burnt once a day for 20 days on the ground level of the cage. The rats were placed on the upper level of the cage to allow maximum exposure to the rising repellent fumes. Liquid mosquito repellent groups were divided into two groups, 3-ml/day and 4ml/day. For 20 days, liquid repellant was sprayed using a nebulizer (Bremmed<sup>®</sup>), a drug delivery device used to administer medication in the form of a mist inhaled into the lungs. Twenty days of exposure in a rat is equal to 18 months in humans [15]. The walls of the cages were covered, and wire mesh was placed at the top to allow for ventilation. After exposure, all groups were maintained under standard conditions until the age of 100 days.

#### 2.3. Histopathology

Prostate glands were removed and fixed in 10% buffered neutral formaldehyde solution for 24 hours at room temperature before being embedded into paraffin blocks. After paraffin embedding, serial of 5 µm thickness sections was then stained. Haematoxylin and eosin (HE) staining method was used to explore luminal epithelial cells, while the Masson's Trichrome staining method was used to explore periductal stromal cells [16]. Six sections and 5 fields per section were randomly chosen in every single rat and examined at magnification ×10 of light microscopy. Histopathological features were examined using a single-blind method by covering the labels at the light microscope level, which an experienced pathologist checked.

HE sections were used to assess the degrees of hyperplasia, cellular polarity (good-bad), epithelial cell polymorphism (negative-positive), chromatin pattern (smooth-rough), and nucleolus presentation (negativepositive) in luminal epithelial cell compartments. The Masson's Trichrome technique differentially stains stromal components and is useful in distinguishing prostate smooth muscle cells (red) from collagen fibers (blue) [17], and was used to assess fibroblast proliferation, smooth muscle arrangement (continuousrare), and prostate gland secretory function.

#### 2.4. Statistical Analysis

The histopathological characteristics of the prostate gland are defined in frequency, and percentage terms, so histopathological characteristics were analyzed as a composite score. The Shapiro-Wilk procedure showed non-normally distributed data, so group comparisons were made using the Mann-Whitney test. In contrast, chi-squared tests were used to assess the differences in distribution frequency between groups. P-value less than or equal to 0.05 were considered significant. The power of this study was 80%, with a 95% confidence interval (CI).

### **3. Results and Discussion**

The histopathological section was focused on the alteration of luminal epithelial and periductal stromal cells of the prostate gland. HE sections used to assess luminal epithelial cells revealed some interesting results regarding insecticide exposure during early life. Among the estrogen group, degrees of epithelial hyperplasia were the highest compared to those of the control and insecticide groups. In contrast, the degrees of hyperplasia in the insecticide groups were

significantly higher than those of the control group. The cellular polarity of epithelial cells was bad in estrogen groups. The highest percentage of bad cellular polarity among insecticide groups was found in the mosquito repellent coil group (27.8%). Epithelial cell polymorphism was found in most subjects of the estrogen group (93%). Surprisingly, polymorphism was not significantly found among the insecticide groups. Chromatin patterns were significantly altered in all treated groups compared to those of the control group: severely rough in all (100%) epithelial cells of the estrogen group; alteration of chromatin pattern was found in 77.7%, 55.5%, and 50% of mosquito coil, 4ml, and 3-ml mosquito liquid repellent, respectively. Nucleolus was found in all subjects (100%) of the estrogen group. Among the insecticide groups, nucleolus was found in 44.4% of the mosquito repellent coil group, in 44% of 4-ml mosquito repellent liquid, and 25% in the 3-ml mosquito repellent liquid group. Luminal epithelial cells were significantly altered in all treated groups compared to those of the control group (see Table 1). The histopathological features of epithelial cell alteration can be seen in Fig. 1.

Table 1 Alteration of a luminal epithelial cell of untreated/control group compared to treated groups (*P*-value  $\leq 0.05$  was considered

significantly different)							
Experimental	Control	Estrogen	Coil	Liquid	Liquid		
Group			repellent	3-ml	4-ml		
Control	-	< 0.001*	< 0.001*	0.002*	0.009*		
Estrogens	< 0.001*	-	< 0.001	< 0.001	< 0.001		
Coil repellent	< 0.001*	< 0.001	-	0.04	0.622		
Liquid 3-ml	0.002*	< 0.001	0.04	-	0.473		
Liquid 4-ml	0.009*	< 0.001	0.622	0.473	-		



Fig. 1 Histopathological features of prostate gland epithelial cell using HE staining method (x100). Hyperplasia of luminal epithelial cells was obviously shown in estrogen [B] (x400), mosquito coil repellent [C], 3-ml [D], and 4-ml mosquito liquid repellent group [E]. Severe chromatin pattern was also observed in the estrogen group [B] (x400).LE=Luminal Epithelial; LEH = luminal epithelial hyperplasia; Cr = chromatin

Many studies have been carried out to seek the effects of estrogenic compound exposure on the prostate gland during pre- and postnatal period [8], [18], [19], [20]. This study used both potent ( $\beta$  estradiol 3-

benzoate) and weak daily used estrogenic compounds, i.e., mosquito repellent coils (transfluthrin 0.03%/coil) and mosquito repellent liquid (transfluthrin 0.162 mg/ml and propoxur 4.05 mg/ml) which caused hyperplasia in the epithelial cells of prostate gland [13], [14].

Prostate formation and differentiation require communication and interaction between epithelial and stromal cells in the prostate. Initially, androgen acts on the mesenchyme to produce a signal for prostate cell stimulation and growth. Subsequently, androgen focuses on epithelial cells for the secretory function of differentiated cells. Reciprocal homeostatic interactions between prostate epithelium and stromal cells are essential for normal prostate growth, development, and function, so disturbing these interactions plays a significant role in carcinoma [21], [22]. In addition to androgens, prostate development is very responsive to estrogenic compounds, especially from environmental EDCs. Estrogenic compound exposure during early life will increase the expression of androgen receptors in the prostate gland and may result in prostate hyperplasia [23]. Studying the effect of EDCs (Bisphenol A) exposure on prostate gland alteration shows some degree of alteration, which includes hyperplasia, inflammatory cell infiltration, and prostatic intraepithelial neoplasia (PIN) [24]. Jarred RA et al. shows that estrogen will alter androgen-regulated prostate development. Thus the evidence reinforces this finding [25].

Cellular alteration was divided histopathologically into three grades depending on the severity of the following changes, i.e., cell crowding and stratification (cellular polarity), nuclear enlargement, cellular polymorphism, chromatin pattern, and nucleolus appearance [26]. The HE sections show that high exposure to potent estrogenic compounds results in severe hyperplasia of luminal epithelial cells with bad cellular polarity, cellular polymorphism, a rough chromatin appearance, and a nucleolar appearance. Therefore, the histopathological characteristic was considered pre-malignant lesions as the main precursor of invasive carcinoma of the prostate, also called PIN [27], [28]. Luminal epithelial cells were significantly altered in all treated groups than untreated groups with P value <0.05. Prostate carcinogenesis involves multiple processes from hyperplasia through dysplasia (pre-cancerous) to carcinoma ranging from low to high histopathological grade, and finally to a cancerous stage (cellular atypia). The HE sections in this study prove that the exposure of estrogen (insecticides) in early life alters the histopathological grading in correlation to the potency of estrogenic compounds.

Insecticide exposure during early life was assessed using the Masson's Trichrome sections, which were particularly focused on periductal stromal cells (fibroblast proliferation, smooth muscle arrangement, and prostate gland secretory function). Fibroblast

proliferation was dominantly found in all (100%) subjects in the estrogen group, 80% of the mosquito repellent coil group (see figure 2), and rarely found in both the 3-ml and 4-ml liquid mosquito repellent groups. Smooth muscle differentiation was found normal (thin and continuous) in the control group. In contrast, the discontinuity of periductal smooth muscle was found in all treated groups, i.e., 100% in both the estrogen and mosquito coil repellent groups, 83.3% in the 4-ml liquid mosquito repellent group, and 60% in the 3-ml liquid mosquito repellent group. Masson's Trichrome stained light blue for normal secretory function and red to none when the secretory function was abnormal or diminished. Alteration of periductal stromal cells was significantly found in all treated groups compared to those of the control group. (see Table 2) The histological features of periductal stromal cell alteration can be seen in figure 2.

Table 2 Alteration of periductal stromal cell untreated/control group compared to those of treated groups (P value  $\leq 0.05$  was considered cignificantly different)

Experimental	Control	Estrogen	Coil	Liquid	Liquid
Group			repellent	3-ml	4-ml
Control	-	< 0.001*	< 0.001*	< 0.05*	0.05*
Estrogen	< 0.001*	-	0.185	< 0.05	0.001
Coil repellent	< 0.001*	0.185	-	0.095	< 0.05
Liquid 3-ml	< 0.05*	< 0.05	0.095	-	0.639
Liquid 4-ml	0.05*	< 0.001	< 0.05	0.639	-



Fig. 2 Histopathological features of prostate gland periductal stromal cell using Masson'sTrichrome staining method (x100). In the control group [A], the smooth muscle was thin and continuous. Smooth muscle bundle and blue reactive stroma were shown in the estrogen group [B] and mosquito coil repellent group [C]. Smooth muscle was decreased and discontinuous in both 3-ml [D] and 4-ml mosquito liquid repellent group [E]. Fibroblast proliferation (blue reactive stromal) was found dominantly in estrogen [A] and coil mosquito repellent group [C]. The secretory function was normal (stained in blue) in the control group [A] and 4-ml mosquito liquid repellent [C], abnormal (stained red) in both mosquito coil repellent [C] and 3-ml mosquito liquid repellent group [D]. Depletion of secretory function was stained none in the estrogen group [A].FB = fibroblast proliferation; SM = smooth muscle were stained red; SMB = smooth muscle bundle; S = secrete

Masson's Trichrome staining method has been used to observe the alteration of periductal stromal cells.

This method can also be used to identify alterations in prostate cancer-reactive stroma (blue reactive/highly collagenous stroma), а modified stromal microenvironment created in response to prostate cancer that accompanies biochemical recurrence and tumor progression [29], [30]. This study shows that neonatal estrogen exposure stimulates the proliferation of periductal fibroblasts, which are clearly visible in the estrogen and mosquito coil repellent groups (See figure 2); additionally, highly collagenous stroma was also identified. Smooth muscle was observed discontinuous in all periductal stroma among treated groups; smooth muscle bundles were found in both the estrogen and mosquito repellent coil groups. Fibroblast proliferation corresponds to a physical barrier that may inhibit branching morphogenesis and interfere with paracrine communication between smooth muscleepithelial interactions in the postnatal prostate. Paracrine communication plays a key role in regulating epithelial differentiation, proliferation and determining the rate of prostate cancer initiation, growth, and progression. Thus, the regulation of cell differentiation will be disrupted [31]. Under the influence of androgen signals towards the prostatic epithelium, prostatic smooth muscles maintain epithelial differentiation and repress epithelial proliferation [32]. In human prostatic adenocarcinoma, loss of smooth muscle cells combined with elevated collagen levels was observed [32], [33]. In the case of cancer, molecular mechanisms of EDCs are in cell cycle regulation and ER-dependent pathways during carcinogenesis [34]. Estrogen exposure during early life will reprogram the prostate gland, causing a permanent alteration in structure and gene expression that leads to an increased susceptibility to prostate lesions or hormonal carcinogenesis by aging [8], [35], [36].

# 4. Conclusion

In conclusion, early exposure to widely-used insecticides that were considered is safe. However, this study proves the disruption of reciprocal communication between periductal stromal and luminal epithelial cells due to fibroblast proliferation and alteration of smooth muscle alignment. Accordingly, it may lead to a predisposition towards neoplastic development in adulthood. Molecular biomarkers of prostate cancer-associated reactive stroma such as thioredoxin reductase 2 protein, the presence of which distinguishes the stroma associated between benign prostatic hyperplasia and prostate cancer, is needed in future studies [37]. Stromal changes might also play a critical role in regulating local cancer growth, invasion, progression, and distant metastasis [38], [39]. Early mosquito insecticide exposure leads to prostate cells differentiation that may predispose to prostate cancer development in later life.

This study did not measure the exact concentration of active ingredients of mosquito insecticides, air transfluthrin concentration inside coil repellent group cage, and air transfluthrin and propoxur concentration inside 3-ml and 4-ml liquid repellent group cages. This is an animal study; thus, generalization regarding humans should be considered with caution.

# 5. Highlights

• Early exposure of pesticides in male rats alters prostate gland cause fibroblast proliferation and smooth muscle alignment

• Early exposure of pesticides in male rats alters reciprocal communication between periductal stromal and luminal epithelial cells, thus, increase the risk of prostate cancer in later life

• Further study needs to be done in animal and human (case-control retrospective cohort design) with the molecular specific marker of prostate cancer

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## References

[1] SHARMA A., MOLLIER J., and BROCKLESBY R. W. K. Endocrine-disrupting Chemicals and Male Reproductive Health. *Reprod Med Biol*, 2020, 19(3): 243-253. <u>https://pubmed.ncbi.nlm.nih.gov/32684823/</u>

[2] REHMAN S., & USMAN Z. Endocrine Disrupting Chemicals and Impact on Male Reproductive Health. *Translational andrology and urology*, 2018, 7(3): 490-503. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6043754/</u>

[3] SCHIFFER C., MÜLLER A., and EGEBERG D. L. Direct Action of Endocrine Disrupting Chemicals on Human Sperm. *EMBO reports*, 2014, 15(7), 758-765. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4196979/

[4] REICHMAN M. E., ALTEKRUSE S., and LI C. I. Feasibility Study for Collection of HER2 Data by National Cancer Institute (NCI) Surveillance, Epidemiology, and End Results (SEER) Program Central Cancer Registries. *Cancer Epidemiol Biomarkers Prev*, 2010, 19(1): 144-147. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2805457/

[5] PRINS G. S. Endocrine Disruptors and Prostate Cancer Risk. *Endocr Relat Cancer*, 2008, 15(3): 649-656. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2822396/</u>

[6] ALAVANJA M. C., SAMANIC C., and DOSEMECI M. Use of Agricultural Pesticides and Prostate Cancer Risk in the Agricultural Health Study Cohort. *Am J Epidemiol*, 2003, 157(9): 800-814.

https://academic.oup.com/aje/article/157/9/800/97345 [7] KOUTROS S., BEANE FREEMAN L. E., and LUBIN J. H. Risk of Total and Aggressive Prostate Cancer and Pesticide Use in the Agricultural Health Study. *Am J*  *Epidemiol*, 2013, 177(1): 59-74. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3590039/</u>

[8] PRINS G. S., HU W. Y., and SHI G. B. Bisphenol A Promotes Human Prostate Stem-Progenitor Cell Self-Renewal and Increases In Vivo Carcinogenesis in Human Prostate Epithelium. *Endocrinology*, 2014, 155(3): 805-817. https://pubmed.ncbi.nlm.nih.gov/24424067/

[9] TARAPORE P., YING J., and OUYANG B. Exposure to Bisphenol A Correlates with Early-Onset Prostate Cancer and Promotes Centrosome Amplification and Anchorage-Independent Growth in Vitro. *PLoS One*, 2014, 9(3): 1-11. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3940879/pd</u> <u>f/pone.0090332.pdf</u>

[10] CALAFAT A., & NEEDHAM L. Human Exposures and Body Burdens of Endocrine-Disrupting Chemicals. *Endocrine-Disrupting Chemicals. Contemporary Endocrinology.* Humana Press, Totowa, USA, 2007: 253-268.

[11] WORLD HEALTH ORGANISATION. State of the science of endocrine disrupting chemicals 2012. WHO Library Cataloguing-in-Publication Data, Geneva, Switzerland, 2013.

https://apps.who.int/iris/handle/10665/78102

[12] RAMOS J. G., VARAYOUD J., and KASS L. Bisphenol A Induces Both Transient and Permanent Histofunctional Alterations of the Hypothalamic-Pituitary-Gonadal Axis in Prenatally Exposed Male Rats. *Endocrinology*, 2003, 144(7): 3206-3215. https://academic.oup.com/endo/article/144/7/3206/2888921

[13] OHTA K., OGAWA T., and SUZUKI T. Novel Estrogen Receptor (ER) Modulators: Carbamate and Thiocarbamate Derivatives with M-Carborane Bisphenol Structure. *Bioorg Med Chem*, 2009, 17(23): 7958-7963. https://www.sciencedirect.com/science/article/abs/pii/S0968 089609009456?via%3Dihub

[14] GO V., GAREY J., and WOLFF M. S. Estrogenic Potential of Certain Pyrethroid Compounds in the MCF-7 Human Breast Carcinoma Cell Line. *Environ Health Perspect*, 1999, 107(3): 173-177. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1566380/

[15] SENGUPTA P. The Laboratory Rat: Relating Its Age With Human's. *Int J Prev Med*, 2013, 4(6): 624-630. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3733029/

[16] O'CONNOR W. N., & VALLE S. A Combination Verhoeff's Elastic and Masson's Trichrome Stain for Routine Histology. *Stain Technol*, 1982, 57(4): 207-210. <u>https://www.tandfonline.com/doi/abs/10.3109/10520298209</u> 066710

[17] TUXHORN J. A., AYALA G. E., and SMITH M. J. Reactive Stroma in Human Prostate Cancer: Induction of Myofibroblast Phenotype and Extracellular Matrix Remodeling. *Clin Cancer Res.* 2002, 8(9): 2912-2923. <u>https://clincancerres.aacrjournals.org/content/clincanres/8/9/</u> 2912.full.pdf

[18] RAMOS J.G., VARAYOUD J., and SONNENSCHEIN C. Prenatal Exposure to Low Doses of Bisphenol A Alters the Eriductal Stroma and Glandular Cell Function in the Rat Ventral Prostate. *Biol Reprod*, 2001, 65(4): 1271-1277. https://academic.oup.com/biolreprod/article/65/4/1271/2723 860

[19] CHANG W. Y., WILSON M. J., and BIRCH L. Neonatal Estrogen Stimulates Proliferation of Periductal Fibroblasts and Alters the Extracellular Matrix Composition in the Rat Prostate. *Endocrinology*, 1999, 140(1): 405-415. https://academic.oup.com/endo/article/140/1/405/2990810

[20] PRINS G. S., TANG W.-Y., and BELMONTE J. Perinatal Exposure to Oestradiol and Bisphenol A Alters the Prostate Epigenome and Increases Susceptibility to Carcinogenesis. *Basic & Clinical Pharmacology & Toxicology*, 2008, 102(2): 134-138. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2819392/pd f/nihms-173883.pdf

[21] MARKER P. C., DONJACOUR A. A., and DAHIYA R. Hormonal, Cellular, and Molecular Control of Prostatic Development. *Developmental Biology*, 2003, 253(2): 165-174. <u>https://core.ac.uk/download/pdf/82134331.pdf</u>

[22] NIETO C. M., RIDER L. C., and CRAMER S. D. Influence of Stromal-Epithelial Interactions on Androgen Action. *Endocrine-Related Cancer*, 2014, 21(4): 147-160. https://erc.bioscientifica.com/view/journals/erc/21/4/T147.x ml?body=pdf-10145

[23] TIMMS B. G., PETERSEN S. L, and SAAL F. S. Prostate Gland Growth During Development is Stimulated in Both Male and Female Rat Fetuses by Intrauterine Proximity to Female Fetuses. *J Urol*, 1999, 161(5): 1694-1701. <u>https://www.auajournals.org/doi/abs/10.1016/S0022-</u>

#### 5347%2805%2969007-6

[24] PRINS G. S, BIRCH L., and TANG W. Y. Developmental Estrogen Exposures Predispose to Prostate Carcinogenesis with Aging. *Reprod Toxicol*, 2007, 23(3): 374-382.

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1927084/pd f/nihms23013.pdf

[25] JARRED R. A., CANCILLA B., and PRINS G. S. Evidence That Estrogens Directly Alter Androgen-Regulated Prostate Development. *Endocrinology*, 2000, 141(9): 3471-3477.

https://academic.oup.com/endo/article/141/9/3471/2989161

[26] HUMPHREY P. A. Grading of Prostatic Carcinoma. *Prostate Pathology*. ASCP Press, Chicago, USA, 2003: 338-74.

[27] HAGGMAN M. J., MACOSKA J. A., and WOJNO K.
J. The Relationship Between Prostatic Intraepithelial Neoplasia and Prostate Cancer: Critical Issues. *The Journal* of Urology, 1997, 158(1): 12-22. <u>https://www.auajournals.org/doi/abs/10.1097/00005392-</u> 199707000-00004

[28] MALHOTRA S., KAZLOUSKAYA V., and ANDRES C. Diagnostic Cellular Abnormalities in Neoplastic and Non-Neoplastic Lesions of the Epidermis: a Morphological and Statistical Study. *Journal of Cutaneous Pathology*, 2013, 40(4): 371-378.

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3809146/pd f/nihms518950.pdf

[29] ELO T. D., VALVE E. M., and SEPPANEN J. A. Stromal Activation Associated with Development of Prostate Cancer in Prostate-Targeted Fibroblast Growth Factor 8b Transgenic Mice. *Neoplasia*, 2010, 12(11): 915-927. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2978914/pd f/neo1211\_0915.pdf

[30] BARRON D. A., & ROWLEY D. R. The Reactive Stroma Microenvironment and Prostate Cancer Progression. *Endocr Relat Cancer*, 2012, 19(6): 187-204. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3716392/pd</u> <u>f/nihms488851.pdf</u>

[31] CHANG W. Y., WILSON M. J., and BIRCH L. Neonatal Estrogen Stimulates Proliferation of Periductal

Fibroblasts and Alters the Extracellular Matrix Composition in the Rat Prostate. *Endocrinology*, 1999, 140(1): 405-415. https://academic.oup.com/endo/article/140/1/405/2990810

[32] CUNHA G. R., HAYWARD S. W., and DAHIYA R. Smooth Muscle-Epithelial Interactions in Normal and Neoplastic Prostatic Development. *Acta Anat (Basel)*, 1996, 155(1): 63-72. <u>https://www.mdpi.com/2077-0383/7/4/68/pdf</u> [33] VELTRI R. W., PARK J., and MILLER M. C. Stromal-Epithelial Measurements of Prostate Cancer in Native Japanese and Japanese-American Men. *Prostate Cancer and Prostatic Diseases*, 2004, 7(3): 232-237. <u>https://www.nature.com/articles/4500738.pdf</u>

[34] YANG O., KIM H. L., and WEON J.I. Endocrine-Disrupting Chemicals: Review of Toxicological Mechanisms Using Molecular Pathway Analysis. J Cancer Prev. 2015 20(1):12-24.

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4384711/pd f/jcp-20-12.pdf

[35] HU W. Y., SHI G. B., and HU D. P. Actions of Estrogens and Endocrine Disrupting Chemicals on Human Prostate Stem/Progenitor Cells and Prostate Cancer Risk. *Mol Cell Endocrinol*, 2012, 354(1-2): 63-73. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3249013/pd f/nihms322813.pdf</u>

[36] PRINS G. S., & HO S. M. Early-Life Estrogens and Prostate Cancer in an Animal Model. *J Dev Orig Health Dis*, 2010, 1(6): 365-370. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4005519/pd</u> f/nihms574151.pdf

[37] SINGER E., LINEHAN J., and BABILONIA G. Stromal Response to Prostate Cancer: Nanotechnology-Based Detection of Thioredoxin-Interacting Protein Partners Distinguishes Prostate Cancer Associated Stroma from That of Benign Prostatic Hyperplasia. *PLoS One*, 2013, 8(6): 1-6. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3675098/pd</u> <u>f/pone.0060562.pdf</u>

[38] KRUSLIN B., ULAMEC M., and TOMAS D. Prostate Cancer Stroma: an Important Factor in Cancer Growth and Progression. *Bosn J Basic Med Sci*, 2015, 15(2): 1-8. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4469930/pd</u> <u>f/BJBMS-15-1.pdf</u>

[39] CHUNG L. W. K., BASEMAN A., and ASSIKIS V. Molecular Insights into Prostate Cancer Progression: the Missing Link of Tumor Microenvironment. *The Journal of Urology*, 2005, 173(1): 10-20. <u>https://www.mdpi.com/2039-4713/11/1/2/htm</u>

## 参考文:

[1] SHARMA A., MOLLIER J., 和 BROCKLESBY R. W. K.

破坏内分泌的化学物质和男性生殖健康。生殖医学生物学, 2020, 19(3): 243-253.
https://pubmed.ncbi.nlm.nih.gov/32684823/

USMAN [2] REHMAN S.. 和 Z. 内分泌于 扰化学物质及其对男性生殖健康的影响。转化性男科和 2018. 7(3): 490-503. 泌尿科. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6043754/ [3] SCHIFFER C., MÜLLER A., 和 EGEBERG D. L. 内分泌干扰化学物质对人类精子的直接作用。欧洲分子 生物学组织的报告, 758-765. 2014, 15(7), https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4196979/

[4] REICHMAN M. E., ALTEKRUSE S., 和 LI C. I. 美国国家癌症研究所监视,流行病学和最终结果计划中 央癌症登记处收集人类表皮生长因子受体2数据的可行性 研究。癌症流行病生物标志物上一个, 2010, 19(1): 144-147.

 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2805457/

 [5]
 PRINS
 G.
 S.

 内分泌干扰物和前列腺癌的风险。内分泌相关癌, 2008,
 15(3):
 649-656.

<u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2822396/</u> [6] ALAVANJA M. C., SAMANIC C., 和 DOSEMECI M. 农业健康研究队列中农业农药的使用和前列腺癌的风险.

美国流行病学杂志, 2003, 157(9): 800-814. <u>https://academic.oup.com/aje/article/157/9/800/97345</u>

[7] KOUTROS S., BEANE FREEMAN L. E., 和 LUBIN J. H.

农业健康研究中的总前列腺癌和侵袭性前列腺癌及农药 使用风险。美国流行病学杂志, 2013, 177(1): 59-74. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3590039/</u>

[8] PRINS G. S., HU W. Y., 和 SHI G. B. 双酚一种促进人类前列腺干祖细胞的自我更新,并增加 人类前列腺上皮体内的致癌作用。内分泌学, 2014, 155(3): 805-817.

https://pubmed.ncbi.nlm.nih.gov/24424067/

[9] TARAPORE P., YING J., 和 OUYANG B. 暴露于双酚一种与早期发作的前列腺癌相关,并促进中心体扩增和体外不依赖锚固的生长. 科学公共图书馆一, 2014, 9(3): 1-11.

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3940879/pd f/pone.0090332.pdf

[10] CALAFAT A., 和 NEEDHAM L. 人与内分泌干扰化学物质的接触和身体负担。破坏内分泌的化学物质。当代内分泌学。人类出版社,美国托托 瓦,2007:253-268.

 [11]
 世界卫生组织.
 内分泌干

 扰化学物质的科学状况,2012年。世界卫生组织图书馆

 出版物编目数据,瑞士日内瓦,
 2013.

https://apps.who.int/iris/handle/10665/78102

[12] RAMOS J. G., VARAYOUD J., 和 KASS L. 双酚一种诱导产前暴露的雄性大鼠下丘脑-垂体-

性腺轴的瞬时和永久组织功能改变。内分泌学, 2003, 144(7): 3206-3215.

<u>https://academic.oup.com/endo/article/144/7/3206/2888921</u> [13] OHTA K., OGAWA T., 和 SUZUKI T. 新型雌激素受体调节剂:具有中号-

碳硼烷双酚结构的氨基甲酸酯和硫代氨基甲酸酯衍生物 。生物有机医学化学, 2009, 17(23): 7958-7963. <u>https://www.sciencedirect.com/science/article/abs/pii/S0968</u> 089609009456?via%3Dihub

[14] GO V., GAREY J., 和 WOLFF M. S. 密歇根州癌症基金会7人乳腺癌细胞系中某些拟除虫菊酯 类化合物的雌激素潜力。环保健康, 1999, 107(3): 173-177.

 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1566380/

 [15]
 SENGUPTA
 P.

 实验老鼠:与人类年龄相关。国际预防医学杂志, 2013,
 4(6):
 624-630.

 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3733029/
 624-630.

27

W. 和 [16] O'CONNOR N., VALLE S. 韦尔霍夫的松紧带和马森的三色染色相结合,用于常规 组织学。染色技术, 1982, 57(4): 207-210. https://www.tandfonline.com/doi/abs/10.3109/10520298209 066710 [17] TUXHORN J. A., AYALA G. E., 和 SMITH M. J. 人类前列腺癌中的反应性基质:肌成纤维细胞表型的诱 导和细胞外基质重塑。临床癌症研究. 2002, 8(9): 2912-2923. https://clincancerres.aacrjournals.org/content/clincanres/8/9/ 2912.full.pdf [18] RAMOS J.G., VARAYOUD J., 和 SONNENSCHEIN C. 产前暴露于低剂量的双酚一种会改变大鼠腹侧前列腺的 围导管间质和腺细胞功能。生物繁殖, 2001, 65(4): 1271-1277. https://academic.oup.com/biolreprod/article/65/4/1271/2723 860 [19] CHANG W. Y., WILSON M. J., 和 BIRCH L. 新生儿雌激素可刺激大鼠前列腺周围膜成纤维细胞的增 殖并改变细胞外基质的组成。内分泌学, 1999, 140(1): 405-415. https://academic.oup.com/endo/article/140/1/405/2990810 [20] PRINS G. S., TANG W.-Y., 和 BELMONTE J. 围产期暴露于雌二醇和双酚一种会改变前列腺表观基因 组并增加对癌发生的敏感性。基础与临床药理与毒理学, 2008. 102(2): 134-138. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2819392/pd f/nihms-173883.pdf [21] MARKER P. C., DONJACOUR A. A., 和 DAHIYA R. 前列腺发育的激素,细胞和分子控制。发展生物学, 2003, 253(2): 165-174. https://core.ac.uk/download/pdf/82134331.pdf [22] NIETO C. M., RIDER L. C., 和 CRAMER S. D. 基质-上皮相互作用对雄激素作用的影响。内分泌相关癌症, 2014. 21(4): 147-160. https://erc.bioscientifica.com/view/journals/erc/21/4/T147.x ml?body=pdf-10145 [23] TIMMS B. G., PETERSEN S. L, 和 SAAL F. S. 子宫内对雌性胎儿的接近度刺激了雄性和雌性大鼠胎儿 在发育过程中前列腺的生长。泌尿外科杂志, 1999. 161(5): 1694-1701. https://www.auajournals.org/doi/abs/10.1016/S0022-<u>5347%2805%2969007-6</u> [24] PRINS G. S, BIRCH L., 和 TANG W. Y. 发育中的雌激素暴露易导致前列腺癌的发生。生殖毒性, 2007. 23(3): 374-382. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1927084/pd f/nihms23013.pdf [25] JARRED R. A., CANCILLA B., 和 PRINS G. S. 雌激素直接改变雄激素调节的前列腺发育的证据。内分 泌学. 2000. 141(9): 3471-3477. https://academic.oup.com/endo/article/141/9/3471/2989161 HUMPHREY P. [26] A. 前列腺癌分级。前列腺病理学。美国临床病理学会出版 社,美国芝加哥,2003:338-74. [27] HAGGMAN M. J., MACOSKA J. A., 和 WOJNO K. J. 前列腺上皮内瘤变与前列腺癌之间的关系:关键问题。 泌尿外科杂志, 1997. 158(1): 12-22.

https://www.auajournals.org/doi/abs/10.1097/00005392-199707000-00004 [28] MALHOTRA S., KAZLOUSKAYA V., 和 ANDRES C. 表皮赘生性和非赘生性病变的诊断性细胞异常:形态学 和统计学研究。皮肤病理学杂志, 2013, 40(4): 371-378. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3809146/pd f/nihms518950.pdf [29] ELO T. D., VALVE E. M., 和 SEPPANEN J. A. 在前列腺靶向成纤维细胞生长因子8b转基因小鼠中与前 列腺癌的发展相关的基质活化。瘤形成, 2010, 12(11): 915-927. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2978914/pd f/neo1211 0915.pdf A.. 和 ROWLEY [30] BARRON D. D. R. 反应性基质微环境与前列腺癌进展。内分泌相关癌, 2012. 19(6): 187-204. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3716392/pd f/nihms488851.pdf [31] CHANG W. Y., WILSON M. J., 和 BIRCH L. 新生儿雌激素可刺激大鼠前列腺周围膜成纤维细胞的增 殖并改变细胞外基质的组成。内分泌学, 1999, 140(1): 405-415. https://academic.oup.com/endo/article/140/1/405/2990810 [32] CUNHA G. R., HAYWARD S. W., 和 DAHIYA R. 正常和肿瘤性前列腺发育中的平滑肌上皮相互作用. 155(1): 巴塞尔解剖学报, 1996, 63-72. https://www.mdpi.com/2077-0383/7/4/68/pdf [33] VELTRI R. W., PARK J., 和 MILLER M. C. 日本本土和日裔男性间质上皮性前列腺癌的测量。前列 腺癌和前列腺疾病, 2004, 232-237. 7(3): https://www.nature.com/articles/4500738.pdf [34] YANG O., KIM H. L., 和 WEON J.I. 破坏内分泌的化学物质:使用分子反应分析的毒理学机 理综述。癌症预防杂志. 2015 20(1):12-24. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4384711/pd f/jcp-20-12.pdf [35] HU W. Y., SHI G. B., 和 HU D. P. 雌激素和破坏内分泌的化学物质对人前列腺干细胞/祖细 胞的作用和前列腺癌的风险。分子细胞内分泌, 2012, 354(1-2): 63-73. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3249013/pd f/nihms322813.pdf PRINS HO [36] G. S., 和 S. M.

[36] PRINS G. S., 和 HO S. M. 动物模型中的早期雌激素和前列腺癌.

健康与疾病的发展起源杂志, 2010, 1(6): 365-370. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4005519/pd</u> <u>f/nihms574151.pdf</u>

[37] SINGER E., LINEHAN J., 和 BABILONIA G. 对前列腺癌的基质反应:基于纳米技术的硫氧还蛋白相 互作用蛋白伴侣的检测将前列腺癌相关基质与良性前列 腺增生相区分。. 科学公共图书馆一, 2013, 8(6): 1-6. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3675098/pd</u> <u>f/pone.0060562.pdf</u> [38] KRUSLIN B., ULAMEC M., 和 TOMAS D. 前列腺癌基质:癌症生长和进展的重要因素。波斯尼亚

基础医学杂志, 2015, 15(2): 1-8. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4469930/pd f/BJBMS-15-1.pdf [39] CHUNG L. W. K., BASEMAN A., 和 ASSIKIS V. 前列腺癌进展的分子洞察:肿瘤微环境的缺失环节。泌

尿外科杂志, 2005, 173(1): 10-20. <u>https://www.mdpi.com/2039-4713/11/1/2/htm</u>